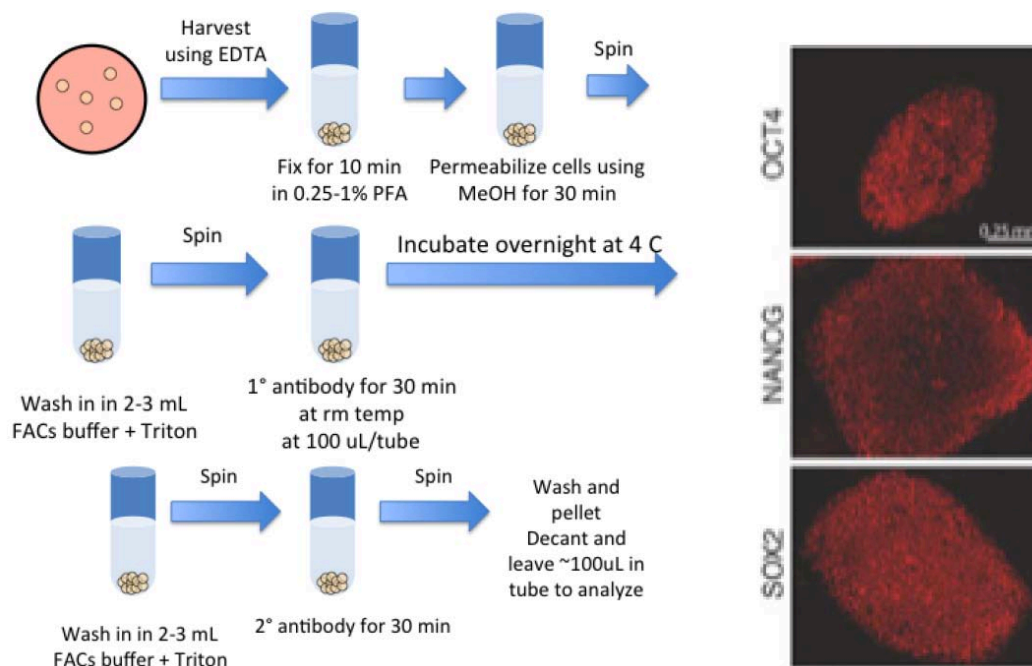


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|------------------|--|
| Title | FACs staining for intracellular protein |
| Date Submitted | May 5, 2012 |
| Submitted by - | Efthymiou, Anastasia - anastasia.efthymiou@nih.gov |
| Adapted from - | Gibco Protocol |
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❖ Introduction:



Staining for intracellular OCT4, NANOG, and SOX2 for FACS analysis¹

❖ Protocol:

A. Harvesting Undifferentiated Cells for FLOW

1. Add 1mL of trypsin/EDTA + 2% chick serum/well of a 6-well plate and incubate for 10 min at 37 C. (*For differentiated cells add 1mL of Dispase for 10 min, remove, add trypsin)
2. Scrape cells and pipette up/down to break up
3. Dilute with FACS buffer and filter the suspension through 100, 80 or 40 micron filter.
4. Spin down cells, 5 min, 1000 rpm
5. Pour off supernatant and add 1mL of PBS
6. Add paraformaldehyde so that the final concentration is 0.25-1% and fix for 10 min in 37 C water bath.
7. Spin down. Resuspend in 2mLs of FACS buffer. (Cells can be placed at 4 degrees for storage if needed)

8. Spin down cells and resuspend in 1mL of ice-cold 90% methanol.
(Alternatively add 100% cold methanol to PBS + fixative to make 90% final concentration)
9. Incubate on ice for 30 min. (Proceed with staining or store cells at -20C in 90% Methanol)
10. Pellet cells.
11. Wash cells by adding 2-3mLs of FACS buffer + Triton. Pellet cells.
12. Pour off supernatant and add 100uL of pre-diluted primary Ab at 1:50.
13. Inc. overnight at 4 C
14. Wash cells by adding 2-3mLs of FACS buffer + Triton. Pellet cells.
15. Pour off supernatant and add 100uL of pre-diluted secondary Ab (1:500).
16. Incubate at RT for 30 min in the dark.
17. Wash and pellet cells. Pour off supernatant to leave ~100ul in tube (Cells can be diluted if needed). Transfer on ice to Flow lab to be analyzed.

❖ **Materials:**

- Begin with 5×10^5 - 1×10^6 cells/tube

| | |
|----------------------------|--------------|
| Trypsin | FACS buffer |
| EDTA | Methanol |
| 2% hick serum | Primary Ab |
| 6-well tissue culture dish | Secondary Ab |
| PBS | Triton |
| 0.25-1% PFA | |

- FACS Buffer

| |
|---------------------------------------|
| PBS (w/o Ca/Mg++) + 2% FBS +0.1% NaN3 |
| *0.5% BSA can be substituted for FBS |

❖ **Troubleshooting:**

❖ **References:**

1. Dirk Hockemeyer, Frank Soldner, et al. Efficient targeting of expressed and silent genes in human ESCs and iPSCs using zinc-finger nucleases. Nat. Biot. 27 851-857 (2009).

Begin

